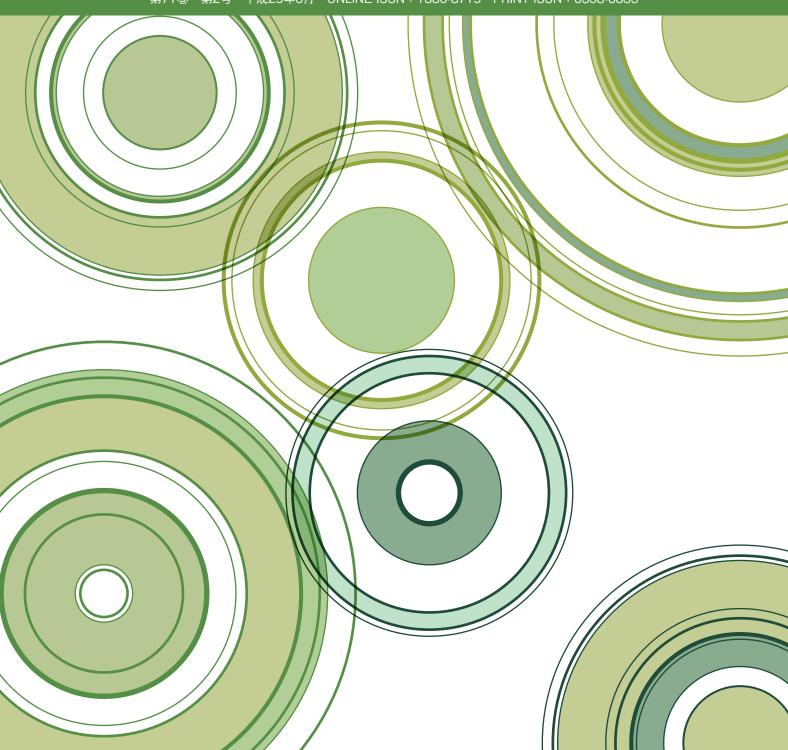
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A testing method examining the dentinogenesis ability of the pulp cells for evaluation of the pulp irritation of the restorative materials

Masamichi Terashita¹, Takahiko Morotomi², Kou Matsuo³, Chiaki Kitamura²

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Abstract

In order to evaluate the pulp irritation by the restorative materials in detail, we developed a new testing method for examining the dentinogenesis ability of the pulp cells in the teeth filled with restorative materials.

Actually, there exist apparent discrepancies between the results of cytotoxicity test and current pulp irritation test with some materials. It led us to develop the new method that enables to evaluate the irritation by restorative materials not by morphological changes but by functional changes of cells for dentinogenesis ability.

The details of this test is as follows: after pulpotomy, the function of the pulp cell recovering from various irritations such as cavity preparation and filling is evaluated in terms of the amount of the formation and the degree of the maturation of dentin bridge formed under the surface of amputated pulp.

Five restorative materials were examined with this method, and the results of the dentin bridge formation and maturation were different among the materials. It indicates that the irritation for the dentinogenesis ability of the pulp cells by the properties of each restorative material can be affected, and the response of pulp cells to the irritations can be quite different from the evaluation with the conventional pulp irritation test. The new pulp irritation test may explain such discrepancies of the result when compared with the cytotoxicity test.

It was confirmed that the new testing method can be useful for pulp irritation test of restorative materials.

Key words: New pulp irritation test / Restorative material / Dentinogenesis ability / Pulpotomy / Dentin bridge

Introduction

The biological evaluations of dental materials and devices in Japan are specified as JIST 0993¹⁾ in accordance with ISO 7405²⁾ and 10993³⁾. Although biological tests for dental materials are generally composed of cytotoxicity test, preclinical test, and clinical test, it is required to carry out *in vivo* simulation test using animal teeth especially for restorative materials for filling teeth.

Histopathological pulp irritation test belonging to the category of mock trial has been established as a method of evaluation for the irritation of restorative materials to dental pulp^{4~16}. This test is composed of some procedures: cavity preparation on vital teeth of animals under general anesthesia, filling with restorative materials of interest, and investigate their pulp responses at each stage after treatment. It is based on the principle that the irritation by materials on the dentin can be directly

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effected to the pulp via dental tubles 17~19).

The pulp irritation test is performed for every newly developed restorative material and many reports have been published 4~16). The irritations by commercial restorative materials have been regarded as slight by the pulp irritation test $^{9\sim16}$. However, there are some reports that these results are markedly different from those of the cytotoxicity test²⁰⁾ and subcutaneous implant test²¹⁾. Especially in resin composites and dental cements, it has been often demonstrated that their irritations of pulp were slight by the pulp irritation test and they would not cause negative influence in clinical situation 9~16) in spite of their considerable cytotoxicities ^{22~28)}. It is, however, easy to assume that irritable constituents in the materials filled in the cavity may pass to the pulp through the dentin of cavity floor 17~19) and they can cause of the non-negligible stimulation of the pulp. In fact, we encounter such cases in which the irritation by material may be suspected to affect the pulpitis in our clinical experiences occasionally.

These discrepancies suggest a possibility that the current pulp irritation test may not fully evaluate the potential of pulp irritation of materials²⁸⁾. As the evaluation criteria of the current test are solely based on the morphological changes of pulp cells^{29~33)}, functional changes of the cells should be revealed by novel test. It is difficult to explain the functional changes of cells from the test merely observing the morphological changes of pulp cells with time. In case that the cell functions decline 34,35, it is well assumed that the various clinical problems may occur. Recently, other symptoms such as allergy have also been pointed out 36,37, and it should be emphasized that the currently approved evaluation methods are not always sufficient for confirming the biological safety of materials.

Therefore, we developed a new method^{38~41)} for detecting functional changes of pulp cells and obtained new findings by examining the pulp irritation by resin composite. This test is composed of pulpotomy after the pulp recovers from various irritations such as cavity preparation and filling, followed by evaluation of the function, dentinogen-

esis ability, of the irritated pulp cells in terms of the amount of the formation and the degree of maturation of dentin bridge under the surface of amputated pulp.

In the present study, we carried out the new test to examine the pulp irritations by five restorative materials and verified the validity of this testing method.

Materials and methods

I. Materials

A. Experimental animals

Twenty-one Wister specific-pathogen-free 9-week-old rats, weight is 250 to 350 g, were used under barrier system condition. They were fed with sterilized water and solid diet and well habituated to the environment before experiment.

B. Restorative materials

The restorative materials used for filling are listed in Table 1. Experimental animals are divided into 5 experimental groups by restorative materials. In order to avoid marginal leakage¹⁴⁾, the restoration site was covered with materials for pit and fissure sealant (Teethmate-A, Kuraray, Tokyo, Japan).

C. Pulp capping agent

Calcium hydroxide (Calvital; CV, Neo, Tokyo, Japan) 42) was used for pulp capping, zinc oxide eugenol cement for lining and amalgam for filling (Table 1).

II. Experimental methods

A. Experimental procedure

The animal protocol was carried out according to the guidelines for animal care of Kyushu Dental University with ethical approval obtained from the institutional panel for animal care.

Rats were deeply anesthetized by intraperitoneal injection of 5% pentobarbital sodium (Nembutal, Sumitomo Dainippon Pharma Co., Osaka, Japan) at a dose of 30 mg/kg, and settled to a specified cavity preparation apparatus^{39,41)} same as reported by Murai⁴³⁾. The experimental teeth and their surrounding area were washed and cleaned with 0.5% chlorhexidine gluconate solution (Hibitane, Sumitomo Dainippon Pharma Co., Osaka, Japan)

Teble 1 N	Iaterials	used
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Code	Material	Product	Manufact	urer	
EZ	Zinc oxide-eugenol cement	Neodyne	Neo	Tokyo	Japan
CR	Resin composite	Clearfil	Kuraray	Tokyo	Japan
		K-echant			
		Clearfil new bond			
AM	Amalgam	Sphrical D	Shoufu	Kyoto	Japan
GI	Glass ionomer cement	Fuji ionomer Type II	G-C	Tokyo	Japan
SC	Silicate cement	Luxsilit	DMG	Hamburg	Germany

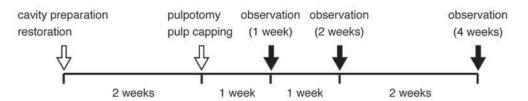


Fig. 1 Experimental procedure

before experiment.

The experimental processes are shown in Fig.1 and Fig.2. Simple cavity was prepared on the mesial aspect of maxillary bilateral first molars at 0.4 mm depth with a #1/2 round bur except for 7 molars as control. After irrigation with 0.5% chlorhexidine gluconate solution and sterilized water and drying, the cavity was filled with one of material, randomly allocated, according to the manufacturer's recommendation. Filled sites were covered with a sealant to avoid marginal leakage, and cusps of lower first molars were ground and removed to prevent occlusion.

Two weeks later, pulpotomy was carried out under deeply anesthesia with above-mentioned. The occlusal surface of pre-treated upper first molar was ground perpendicularly from above its central cusp to a depth of 1.4 mm to form the standardized amputation with a #1/2 round carbide bur on the specified cavity preparation apparatus^{39,43)}. After the surface of amputated pulp was washed alternately with 10% hypochlorite solution and 3% hydrogen peroxide, subsequently washed with physiological saline and dried, amputated pulp was capped with calcium hydroxide, lined with zinc oxide eugenol cement, and filled with amalgam.

As a control group, the same pulpotomy was

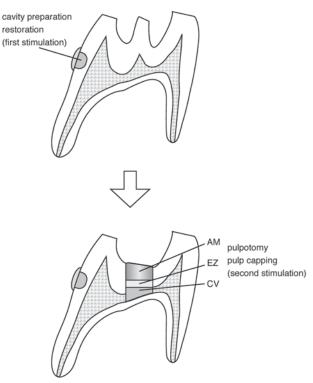


Fig. 2 Schematic illustration of the experimental tooth

performed for intact teeth without any treatment. The test was carried out for 7 teeth for each of experimental and control groups.

B. Histological observation

The postoperative intervals of sacrifice were 1, 2,

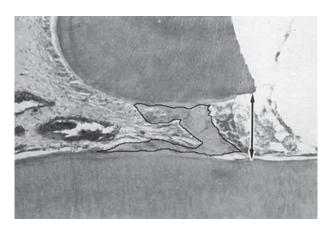


Fig. 3 Thickness of dentin bridge were examined below. The thickness of dentin bridge (μm) = Area of dentin bridge (□)/Diameter of amputated pulp (\$\$\frac{1}{2}\$)

and 4 weeks after pulpotomy. After each interval, rats were euthanatized by excessive dose of 5% pentobarbital sodium. Immediately maxillary segments including the experimental teeth were extracted and immersed in 10% neutral formalin solution. After completely fixation, they were decalcified in Plank and Rychlo rapid decalcifying solution for 5 days, neutralized by 5% sodium sulfate solution, washed by water and dehydrated by graded ethanol. Then the teeth were paraffinembedded following common procedures and were serially cut at a thickness of 5 μ m. Hematoxylin and eosin staining and Hucker-Conn Gram staining were performed for the specimens.

III. Method of evaluation

Five specimens from each tooth were analyzed. The thickness and maturation level of the dentin bridge formed under the surface of amputated pulp and the morphology of the pulp just below the dentin bridge were obserbed.

The thickness of the dentin bridge was calculated from the following formula and the data were averaged (Fig. 3).

Thickness of dentin bridge $(\mu m) = Area$ of dentin bridge/Diameter of amputated pulp

NIH image (National Institutes of Health, Bethesda, MD) was used for measurement and the data were statistically analyzed by one way ANOVA test.

The maturation level of the dentin bridge was

expressed as (+) for highly calcified dentin having tubular structure, (-) for osteodentin having no tubular structure and (\pm) for mixture of both dentins.

Results

Table 2 and Fig. 4 show the changes in thickness with time of the dentin bridge formed under the surface of amputated pulp of the tooth filled with restorative material at the mesial side before pulpotomy. Histopathological features of the surface area of amputated pulp in the filled teeth at 1 week and 4 weeks are exhibited in Fig. 5 to Fig.10, and the degrees of maturation of the dentin bridge at 4 weeks are shown in Table 2.

In the teeth filled with zinc oxide eugenol cement (EZ-P), the thickness of the dentin bridge and its increments with time were almost equivalent to those in the unfilled teeth (NO-P) through all the experimental periods. The teeth filled with resin composite (CR-P) tended to show smaller thickness of the dentin bridge than the teeth filled with other materials at every period, and the thickness was significantly smaller than NO-P at 1 and 2 weeks (p < 0.01). In the teeth filled with amalgam (AM-P), the thickness increased in an approximately equivalent ratio from 1 week through 4 weeks and it showed significantly smaller thickness than the unfilled teeth at 1 and 2 weeks (p<0.05, 0.01). Marked increases in thickness of the dentin bridge were found from 1 week to 2 weeks in the teeth filled with glass ionomer cement (GI-P) and silicate cement (SC-P). The GI-P was obserbed larger thickness than the NO-P at 2 and 4 weeks (p<0.05, 0.01). In the SC-P, the thickness was smaller at 1 week (p<0.05) and larger at both 2 and 4 weeks (p<0.05, 0.01) than in the NO-P.

The NO-P group at 1 week was showed a formation of poorly calcified dentin bridge with cellular dentin beneath the necrotic zone. At 4 weeks a highly calcified dentin bridge with tubular structure and palisading of spindle cells was observed (Fig.5). The EZ-P group at 1 week showed an early-formed dentin bridge consisting of

Experimental	Average thickness (μm)				
group	1 week	2 weeks	4 weeks	_ mat	

Teble 2 Evaluation of dentin bridge

Experimental	Average thickness (μm)			Degree of
group	1 week	2 weeks	4 weeks	maturation
NO-P	105 ± 27	192 ± 25	216 ± 29	+
EZ-P	87 ± 21	164 ± 38	194 ± 17	+
CR-P	$24\pm14**$	$129\pm17^{**}$	$187\!\pm\!28$	+
AM-P	$51 \pm 39^*$	$132 \pm 32**$	232 ± 27	\pm
GI-P	$85\!\pm\!27$	$228\!\pm\!26^*$	$274 \pm 30**$	\pm
SC-P	$54 \pm 35*$	$242 \pm 40*$	316±51**	_

*P<0.05 **P<0.01

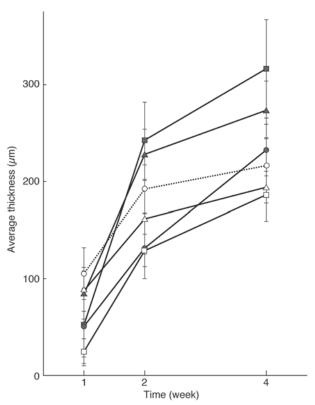
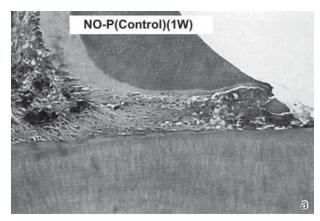


Fig. 4 Changes in thickness with time of the dentin bridge formed on the surface of amputated pulp. O··· NO-P (Control) —△— EZ-P -**●**— AM-P **−▲**— GI-P - CR-P

basophilic demarcation and acidophilic matrix layers beneath the necrotic zone. At 4 weeks, a formation of highly calcified dentin bridge with tubular structure and odontoblast-like cells just below the dentin bridge was observed (Fig.6). These features were almost the same as those in the NO-P group (Fig.5). The CR-P group at 1 week showed an incomplete closure of the wound by an eosinophilic matrix beneath the necrotic zone. At 4 weeks, similar to those in control group, highly calcified dentin bridge with tubular structure was formed (Fig.7). The AM-P group at 1 week showed an incomplete closure of the wound by a dentin bridge with tubular structure and palisading of cuboid odontoblasts. At 4 weeks, highly calcified dentin bridge with scarcely any tubular structure and palisading of oval-shaped odontoblasts just below the dentin bridge was observed (Fig.8). The GI-P group at 1 week showed an incomplete closure of the wound by a dentin bridge consisting of largely of tubular dentin as well as a cellular dentin area just below the necrotic zone. At 4 weeks, a continuous formation of both a cellular dentin bridge and a highly calcified tubular dentin bridge was observed with palisading of cuboid odontoblasts just below them (Fig.9). The SC-P group at 1 week showed a closure of wound by eosinophilic matrix beneath the necrotic zone and palisading of ovoid cells just below the eosinophilic matrix. At 4 weeks, a formation of dentin bridge consisting of both a cellular and vascular area and a nearly acellular osteodentin-like area was observed (Fig.10).

In the teeth filled with restorative materials, pulp hyperemia was observed adjacent to the dentin bridge in all of the specimens, although recovery from the circulatory disturbance of the pulp just below the dentin bridge was noted in the unfilled control specimens.

In addition, only a little penetration of bacteria was recognized in a few cases, which was limited to a part of the cavity wall and no changes in pulp



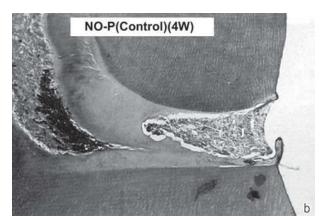
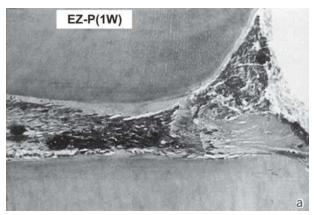


Fig. 5a NO-P (Control) group, near the wound pulpotomy, 1-week postpulpotomy (H·E., ×50). Beneath the necrotic zone, a formation of poorly calcified dentin bridge with embedded cells is observed. Hyperemia without inflammatory cell infiltration is noted in the adjacent pulp tissue.

b NO-P group, near the wound pulpotomy, 4-week postpulpotomy (H·E., ×50). A highly calcified dentin bridge with tubular structure is shown. Palisading of spindle cells just below the dentin bridge is noted. The adjacent pulp tissue recovers the circulatory impairment.



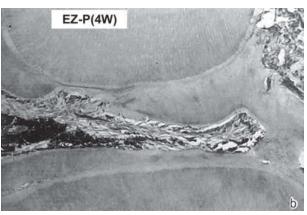


Fig. 6a EZ-P group, near the wound pulpotomy, 1-week postpulpotomy (H·E., ×50). Beneath the necrotic zone, an early-formed dentin bridge (87 μm) consisting of basophilic demarcation and acidophilic matrix layers is noted. Hyperemia is observed in the neighboring pulp tissue.

b EZ-P group, near the wound pulpotomy, 4-week postpulpotomy (H·E., ×50). This shows complete closure of the wound by highly calcified dentin bridge with tubular structure and spindle cells just below the dentin bridge. Hyperemia is noted in the pulp.

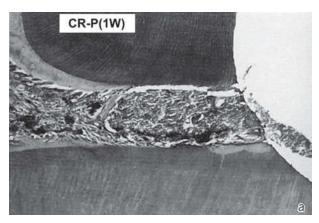
response were detected by such an existence of bacteria.

Discussion

Actually, there exist apparent discrepancies between the results of cytotoxicity test and current pulp irritation test in some materials $^{4\sim16,20\sim28)}$. It led us to the idea of the present study to evaluate the irritation of restorative materials not by morphological changes but by functional, dentinogenesis ability, changes of cells $^{38\sim41)}$.

It has so far been demonstrated, for example,

that the pulp disturbance in the teeth filled with resin composites would be caused not by the irritation by resin composites themselves but by bacteria remaining in the cavity or penetrating through marginal gaps^{14~16)}. It was, therefore, once discussed that resin composites might be available for a direct pulp capping material^{44~46)}. However, cytotoxicity was clearly recognized in resin composites^{22~27)}, even though it showed good results in pulp irritation test^{10~16)}. Furthermore, extensive apoptosis of the pulp cell was observed in the filling with resin materials according to the



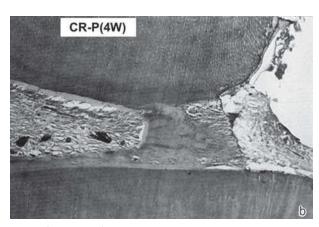
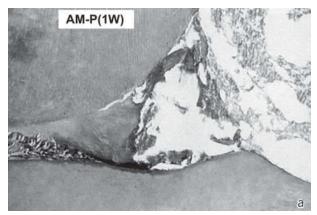


Fig. 7a CR-P group, near the wound pulpotomy, 1-week postpulpotomy (H·E., ×50). Beneath the necrotic zone, an incomplete closure of the wound by an eosinophilic (acidophilic) matrix was observed. Hyperemia was noted in the adjacent pulp tissue.

b CR-P group, near the wound pulpotomy, 4-week postpulpotomy (H·E., ×50). This shows highly calcified dentin bridge formation with tubular structure and palisading spindle cells just below the dentin bridge. Mild hyperemia is noted in the adjacent pulp tissue.



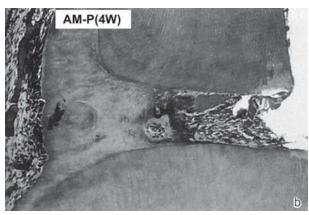


Fig. 8a AM-P group, near the wound pulpotomy, 1-week postpulpotomy (H·E., ×50). Beneath the necrotic zone, an incomplete closure of the wound by a dentin bridge with tubular structure is observed. Palisading of cuboid odontoblasts just below the dentin bridge is noted. The adjacent pulp tissue shows no remarkable changes.

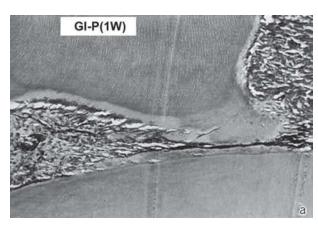
b AM-P group, near the wound pulpotomy, 4-week postpulpotomy (H·E., ×50). Beneath the necrotic zone, a highly calcified dentin bridge with almost no tubular structure is observed. Palisading of oval-shaped odontoblasts just below the dentin bridge is noted. The adjacent pulp tissue shows hyperemia.

conventional pulp irritation test method⁴⁷⁾. It was detected that the apoptotic cells were not limited to the pulp just below the cavity but widely spread over the entire pulp.

Cytotoxicity test is apt to be unvalued because it is much apart from the clinical situation compared with the pulp irritation test taking account of the existence of dentin. However, dentin is tubular structure and it is anticipated that the irritable constituents dissolved from material can easily penetrate to the pulp $^{17\sim19}$, and cause pulp disorder or sometimes bring about pulp necrosis by

following combined irritations. The current pulp irritation test is unable to identify such possibility of risks. We also have no bases for above speculations and it is necessary to confirm them by some other investigations. Under these circumstances, the pulp response to restorative materials was examined utilizing the new testing method in the present study.

The new method aims to evaluate whether the pulp irritated by restorative materials in the cavity may maintain its original reparative function or not in terms of amount of formation and degree of



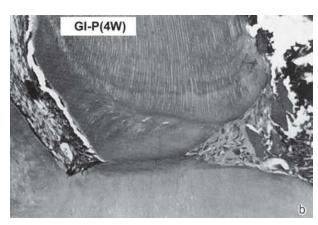
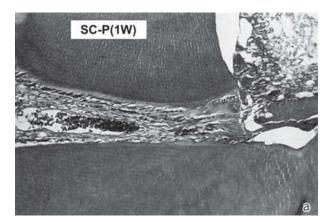


Fig. 9a GI-P group, near the wound pulpotomy, 1-week postpulpotomy (H·E., ×50). An incomplete closure of the wound by a dentin bridge consisting largely of tubular dentin is observed, though a cellular dentin area is also noted just below the necrotic zone. The adjacent pulp tissue shows no remarkable changes.

b GI-P group, near the wound pulpotomy, 4-week postpulpotomy (H·E., ×50). This shows a formation of dentin bridge consisting of two contiguous, but distinct dentin: a cellular dentin-like tissue and a highly calcified tubular dentin. Palisading of cuboid odontoblasts is observed just below the dentin bridge. Mild hyperemia is noted in the adjacent pulp tissue.



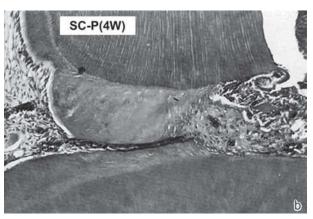


Fig. 10a SC-P group, near the wound pulpotomy, 1-week postpulpotomy (H·E., ×50). Beneath the necrotic zone, this shows a closure of wound by eosinophilic (acidophilic) matrix just below which palisading of ovoid cells is observed. Hyperemia with markedly dilated blood vessels is noted in the adjacent pulp tissue.

b SC-P group, near the wound pulpotomy, 4-week postpulpotomy (H·E., ×50). Beneath the necrotic zone, this shows a formation of dentin bridge consisting of two continuous areas: a cellular and vascular area and a nearly acelluar osteodentin -like area. Mild hyperemia is noted in the adjacent pulp tissue.

maturation of dentin bridge. In this method, the formation of the dentin bridge was observed following pulpotomy from other tooth surface at the time of the pulp has recovered from the response to irritations by several filling procedures ^{8,34)}. By comparing the results among the restorative materials, it is assumed to be able to evaluate the reparative function, dentinogenesis ability, of the pulp cells subjected to the irritation of restoration. It may also be possible to observe the effects to the pulp at the far site of pulpotomy region and appearance of reparative dentin, and perhaps this method

will allows for the comprehensive evaluation³⁹⁾. Furthermore, it may be possible to perform the pulp examination for pulp capping agents if the pulp-exposed cavity is provided⁴¹⁾.

The present results showed that there were significant differences in amount of formation and degree of maturation of the dentin bridge among the used restorative materials, which apparently indicates that the irritation by restorative materials affect the reparative function of the pulp cells. It is also indicated that the irritation by restorative materials is not limited to the pulp just below the

cavity but spread over the entire pulp.

The formation of the dentin bridge in the pulp subjected to the irritation by resin composite was delayed and that in the pulp subjected to the irritations by silicate cement or glass ionomer cement of which liquid components were acid was accelerated compared with the pulp of unfilled tooth as the control. The amount of formation of the dentin bridge with amalgam linearly increased and that with zinc oxide eugenol cement showed almost equivalent level to the control. The degrees of maturation of the dentin bridge in the pulp subjected to the irritations by zinc oxide eugenol cement and resin composite at 4 weeks were similar to that of the control, those of amalgam and glass ionomer cement were slightly inferior and that of silicate cement was apparently inferior to the control.

Based on the newly developed pulp irritation test, the irritations by resin composite, amalgam, glass ionomer cement, and silicate cement affect the reparative function of the pulp, although the morphological changes of the pulp cells seem to be reversible^{29~33)}. The results suggest that the irritation of resin composite decreased the dentinogenesis ability of odontoblasts and that of silicate cement, on the other hand, delayed the differentiation of odontoblasts, and consequently mature dentin bridge formation was suppressed in spite of the abundant formation of immature dentin bridge. Nakayama et al. reported from an examination by the same evaluation method as this study that when resin composite was filled on the exposed pulp in the prepared cavity, the dentin bridge was abundantly formed on the surface of the amputated pulp, even though the dentin was immature⁴¹⁾. The results were not similar to those for resin composite but rather similar for silicate cement in this study. The difference in the pulp response to resin composite between them may be arisen by the existence of the cavity floor dentin. In this study the permeability of its constituents to the pulp would be decreased by the cavity floor dentin, whereas it was filled on the exposed pulp with bonding agent and its

constituents were directly in contact with the pulp in the test of Nakayama et al.⁴¹⁾. Anyway, both of the resin composite and silicate cement may have a high risk of disturbing the reparative ability of the pulp cells even in the clinical use if the cavity floor dentin cannot completely intercept the passage of all the irritants from the restorative materials. As for amalgam and glass ionomer cement, complex mixtures of mature and immature dentin bridges were found. It suggests that their irritations would cause further complicated pulp responses. The effects of amalgam on reparative function of the pulp cells are somewhat strong since the decreased dentinogenesis ability, and those of glass ionomer cement are relatively slight.

As for zinc oxide eugenol cement which had been judged to be irritable by the cytotoxicity test²⁰⁾ caused no pulp disturbance because of no passage of constituents to the pulp when it was filled in the cavity, and it was confirmed to be available for a control^{48, 49)} in pulp irritation test.

Conclusion

In order to evaluate the pulp irritation by the restorative materials in detail, we developed a new testing method for examining the dentinogenesis ability, amount of formation and degree of maturation of dentin bridge, of the pulp cells in the teeth filled with restorative material.

Five restorative materials were examined with this method, and it was observed that the amount of formation and degree of maturation of the dentin bridge were different among the materials. It indicates that the irritation of dentinogenesis ability of the pulp cells by the restorative material is distinctive of each material, which is quite different from the evaluation with the conventional pulp irritation test.

It was confirmed that the new testing method is useful for pulp irritation test by restorative materials, and thus the test may be able to explain such discrepancies of the result when compared with the cytotoxicity test.

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歯髄細胞の象牙質形成能を指標とする修復材料の歯髄刺激試験

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抄 録

修復材料の歯髄刺激を評価するために歯髄細胞の機能(象牙質形成能)を指標とした新しい試験方法を開発した. 現在行われている歯髄試験は修復材料を窩洞に充塡後、刺激を受けた歯髄細胞の形態変化を観察することで評価を行っている.この評価法では、各修復材料間の歯髄刺激に差はほとんどなく、細胞毒性試験との乖離がみられる.そこで、歯髄細胞の形態変化では無く、機能の変化に着目して評価することにした.

新しく開発した試験方法は、各修復材料を充塡した歯の歯髄が窩洞形成や充塡による種々の刺激から回復した後、他の部位から歯髄断髄法を行い、断髄面に形成されるデンティン・ブリッジの量や成熟度で修復材料の歯髄刺激を評価する方法である。今回、新しい歯髄試験を用いて、5種類の修復材料の歯髄刺激を評価するとともに、この試験の有用性を検討した。

新しい歯髄試験では、各修復材料間で象牙質形成能に特徴的な違いが認められ、差の認められなかった従来の歯髄 試験とは異なった結果となった。これらの結果は細胞毒性試験との矛盾を説明することも可能であった。新しい歯髄 試験は修復材料の歯髄刺激を評価するために有用であることが示された。

キーワード:新規歯髄試験/修復材料/象牙質形成能/歯髄断髄法/デンティン・ブリッジ

ポーセレン・ラミネートベニア修復の25年経過症例

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A Case Report of a 25-year Clinical Follow-up of Porcelain Laminate Veneer Restoration

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Abstract

Porcelain laminate veneer (PLV) restoration is generally accepted as one of aesthetic dental treatment to improve the discoloration of teeth. However, few long-term clinical studies have been reported. The present report shows a 25-year clinical follow-up case of PLV restorations that were carried out to improve severe discoloration of the maxillary anterior teeth (321|123) by tetracycline.

Nine years after the first restorative treatment, partial fractures around cervical area of PLV on <u>23</u> were observed. These fractured PLV of <u>23</u> were removed, and re-restored by new PLVs. Twenty-five years after the first restoration, all PLVs were re-treatment because of the color change.

Key words: Porcelain laminate veneer restoration / Long-term observation / Cervical fracture / Re-treatment

抄 録

歯の変色による審美障害に対する治療法の一つとしてポーセレン・ラミネートベニア(PLV)修復があるが、長期経過報告はほとんどない. 今回我々は、テトラサイクリンによる重度の変色歯で上顎前歯部の審美障害を主訴とする患者にPLV修復を行った症例で25年という長期経過を観察できたので報告する.

この症例は9年経過後に123の歯頸部にPLVの部分破折が生じ、2歯のみ再修復を行い、25年経過後に明度が低下

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したという患者の訴えのもとに全てのPLV(321|123)を再修復したものである.

キーワード:ポーセレン・ラミネートベニア修復/長期経過症例/歯頸部の部分破折/再修復

緒 言

ポーセレン・ラミネートベニア (Porcelain Laminate Veneer; PLV) 修復は、変色歯や正中離開・矮小歯等の形態不全歯の改善のための治療法の一つとして開発¹⁾・応用されてきた術式である。九州歯科大学口腔保存治療学分野では、27年前に最初の症例を報告し²⁾、基礎研究とともに臨床術式の確立を行ってきた^{3~10)}.これは歯質削除量が少なく、歯への侵襲を最小限にして審美性を回復させる比較的新しい術式であり、長期症例の報告は少ない¹¹⁾.当分野では3年前に15~18年の間良好に経過した3症例を報告した¹²⁾.今回は25年というさらに長い期間観察でき、観察期間中に一部のベニアの破折による再修復と25年経過後に明度の低下という訴えに対して全てのPLVを再修復した症例について報告する.

症 例

初診:1988年6月29日

患 者:16歳,女性.

主 訴:前歯部の審美障害(歯の変色)

既往歴:乳児期に母親が腎盂炎にてテトラサイクリン系

抗生物質を服用,他に特記事項無し.

家族歴:特記事項無し.

現病歴:変色歯の治療を希望して近医を受診. PLV修復の適応であると判断され,治療目的で当院保存治療科を紹介.

現 症:口腔内所見;歯面は歯頸部側に帯状の濃い灰褐 色を呈した変色が認められる. とくに上顎6前歯の変色 が強い. 歯列は軽度の叢生を示す. 歯間乳頭部歯肉に軽度の発赤と腫脹が認められる(図1-A). 上顎前歯は歯髄電気診に正常に反応し, エックス線写真(上顎前歯部)では, 特記すべき異常所見は認められない(図1-B).

診 断:歯頸部に帯状の変色を伴う永久歯の石灰化不全症(新Feinmanの分類Ⅲ)

治療方針: PLV修復

処置および経過: 初診日よりブラッシング指導と Professional Tooth Cleaning (PTC) およびスケーリングを行い、プラークコントロールによる歯肉の改善を 図った. 1988年8月2日に321/123の支台歯形成 (切縁非被覆型) $^{2)}$ を行い、1週間後に通法に従いPLVを装着した (図 1 -C).

PLVのシェードはA2とし、コスモティック・ポーセレン(ジーシー社製)を耐火模型上で築盛・焼成した、変色した歯の色を遮断するためマスキングデンティン・ポーセレンを使用した、PLV完成後、コスモティック・セメント(ジーシー社製)のUniversal色を用いて装着した。

術後の患者評価:治療して良かったと満足している.

リコール1:1993年1月7日

装着4年5か月経過後の口腔内写真を示す(図2). や や明度が低下しているが、PLVに問題はない. 歯髄電気 診にも正常に反応し、臨床的な問題は認められない.

リコール2:1997年8月19日(部分破折への再修復) 装着後9年経過時の口腔内写真を示す(図3-A). 上顎左側側切歯および犬歯の歯頸部にPLVの部分破折











図1 初診時(1988年)

A: 術前の口腔内写真 B: 術前のレントゲン写真

C:PLV装着直後



図2 リコール 1 (1993年) 修復後4年5か月

が認められた. 乳頭部歯肉に軽度の発赤と腫脹が認めら れる. 歯髄電気診に正常に反応し、他に異常所見は認め られない.

処置および経過: 23 の部分破折したPLVを全て除去し、 支台歯形態の修正(切縁非被覆型)を行い(図3-B), 1 週間後にPLVを装着した(図3-C). この時<u>23</u>の歯頸 部に1~2mm程度象牙質の露出を認めたため、象牙質 部においてはエッチングに加えデンティンプライマー処 理を行った.

PLVのシェードはA3とし、A3に相当するコスモ ティック・ポーセレンとマスキングデンティン・ポーセ レンを使用した. 患歯へのPLV装着はコスモティック・ セメントのBrown色を用いた.

再初診:2013年11月1日(再修復)

主 訴:治療した前歯が変色してきたのでより白い PLVの再製を希望

治療方針: PLV再修復

現 症:装着後25年経過時の口腔内写真を示す(図 4-A). 3」歯頸部に若干の歯肉退縮, および<u>[23</u>間乳頭 部歯肉に軽度の発赤と腫脹が認められる.歯髄電気診に 正常に反応し、エックス線所見でも異常は認められない (図4-B). 他に臨床的異常所見も認められない. 他覚 的所見として、PLVの色調は修復時より明度がやや低下 しているように思われる.

処置および経過:ブラッシング指導およびPTCを行っ た後、2014年1月14日に321 123 PLVを全て除去し、再 度支台歯の形態修正(切縁非被覆型)を行い(写真5-A). 1週間後に通法に従いPLVを装着した(図5-B). 初診 時より歯列の叢生を認め、上顎右側犬歯はやや唇側に転 位している. 歯肉退縮により歯根面が露出しているが, スマイルラインでは歯根露出部が見えないことと生活歯 であり切削量を最小限に留めるという点から、接着で保 持するPLVの術式の原則^{2, 7, 13)}に従い, 今回の再修復 に際しては根面被覆せず窩洞形成はエナメル質内に留め

PLVのシェードはA2とし、Super Porcelain AAA







図3 リコール2(1997年):修復後9年

A:PLVの部分破折

B: 支台歯形態修正および歯面処理後

C:PLV装着直後











図4 再初診時(2013年):修復後25年

A:口腔内写真 B:レントゲン写真



B

図5 再修復(2014年) A:支台歯形態修正後 B:PLV再修復直後



図6 リコール(2014年) 再修復後7か月

(Noritake社製)を耐火模型上で築盛・焼成した。また、初診時同様に歯の色を遮断するためマスキングデンティン・ポーセレンを使用した。PLV内面には通法通りサンドブラスター処理に加えシランカップリング処理を行い、歯面はエッチング処理と象牙質露出部にはデンティンプライマー処理を行った。より白くしたいという患者の希望を考慮し、クリアフィル・エステティックセメント(Kuraray社製)のブリーチ色を用いてPLVを装着した。

術後の患者評価:再治療をして良かったと満足している.

リコール:2014年8月14日

装着後7か月経過時の口腔内写真を示す(図6). PLV に問題はなく歯髄電気診に正常に反応し、臨床的な問題は認められない. 一部の辺縁歯肉に軽度の発赤を認めるが、再初診時と同程度である.

考察

本症例は、重度のテトラサイクリン変色歯による審美障害に対してPLV修復を応用し、安定した長期経過が得られた1症例である。接着を応用した術式での20年を超える長期症例の報告¹³⁾はほとんどない。今回、25年という長期症例が観察できたので若干の考察を加え報告する。

25年という長い間には、部分的な歯肉退縮による歯 頸部歯根面の露出やPLVの明度の低下、PLVの一部破 折等が見られたが、歯周組織や歯髄に臨床的問題はなく 経過はおおむね良好であったと言える。患者には初診時 よりセルフケアであるプラークコントロール法を指導 し、清掃状況も良好であった。

この術式は審美歯科処置の一つであり露出する変色し た唇頬側歯面を全て覆う必要があるため、本来であれば 歯肉の位置が安定する20歳以降が適応症である¹²⁾. 加藤 ら (1999) 14) によると、PLV修復10 ~ 11年間の臨床成績 の結果, 生理的歯肉退縮傾向が少なくなる25~28歳ぐ らいに実施した方がより長期にわたって安定した臨床成 績が得られると推察している. 本症例は初診時が16歳 であったが、社会的・心理的背景を十分に情報収集・分 析しインフォームドコンセントを行った結果、患者の QOL向上のためPLV修復を選択した. 歯肉退縮に関し ては、PLV装着から9年経過後(25歳当時)でも歯頸部 歯面の露出は認められなかった。その後の16年間で認 めた歯肉退縮は、唇側転位している1歯(犬歯)に限局し た最大で約1.5mmの退縮であった. 術前の状態から考え ると歯周疾患等の病的因子に起因するものである可能性 は低いと考えられ、生理的な範囲の歯肉退縮量と判断で きる15~17). 今後も適切なセルフケアを続けることが必 要である.

この症例では9年後に2歯(|23)のPLVが歯頸部付近で破折し、一部が脱離した。脱落の原因として、装着時に固定の不備によりセメントが厚くなったこと(カルテに記録有り)と、この部位の象牙質露出による接着が弱くなったことが挙げられる。咬合力によってPLVの歯頸部に強い引っ張り応力が発生し、相対的に強度の小さいセメント内にはPLVを引き剥がす方向にかなりの応力が生じる⁸⁾。歯質の削除をエナメル質内にとどめるという支台歯形態^{1,2,5,7)}を遵守することが重要であるが、

象牙質が露出した場合はデンティンプライマー処理を行い,確実な接着操作でセメントを薄くすることが破折や 脱離を防ぐことにつながる.

今回、PLVの明度の低下に関してはPLV自体の変色ではなく、長期にわたるレジンセメントの色と透過性の変化による影響が推察¹²⁾されたが、明確な原因に関しては不明である。最近ではレジンセメントの性質も格段に向上しており、吸水性も改善され経時的な変色も少ないことが期待される。今後も定期的な経過観察を行っていく必要がある。PLV修復は修復法の中で最も寿命の長い修復法に属し^{12, 18)}、しかも歯の保存が図れ、生活歯の変色や軽度の形態不全を改善する最も推奨される術式であるといえる。

まとめ

今回我々は、重度のテトラサイクリン変色歯による審美障害に対して行ったポーセレン・ラミネートベニア修復治療後の経過と9年後に歯頸部の破折により2歯を再修復、25年後に明度の低下という訴えにより全歯(上顎6前歯)を再修復した長期症例について若干の考察を加えて報告した。本症例を通して、PLV修復は長期にわたって色の明度がやや低下してもPLV再製可能な歯質保存ならびにふさわしい色調の再現が可能という審美性の観点から、生活歯の変色に対して極めて有効な術式であることが示唆された。

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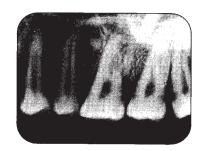
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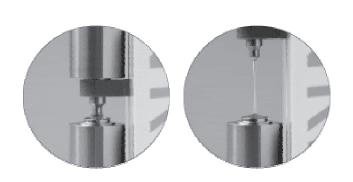






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